

Impact of Micro-Aid[®] on the Bacterium Salmonella¹

Introduction

Bacteria belonging to the genus *Salmonella* are rod-shaped (bacillus), gram-negative, facultative anaerobic bacteria in the family *Enterobacteriaceae*. Based on their phenotypic profile, the genus *Salmonella* is divided into two species, *S. enterica* and *S. bongori*. *Salmonella enterica* is further divided into six subspecies that include over 2,500 serotypes. The majority (1,531) of these serotypes belong to Salmonella *enterica* subsp. *enterica* (I) and were originally given names such as Typhimurium, Dublin, and Infantis. *Salmonella enterica* subspecies *enterica* (I) is the only subspecies that has been associated with disease in warm-blooded animals and humans. It is estimated that 99.5% of *Salmonella* strains isolated from humans and warm-blooded animals belong to this subspecies. This gives rise to a food safety issue for animal agriculture industries.

Materials and Methods

Six serotypes of *Salmonella* were tested for their susceptibility to Micro-Aid[®] using the agar dilution test as described in the Clinical and Laboratory Standards Institute (CLSI) M31 document. The six different serotypes of *Salmonella* were *Salmonella* serotype (ser) Infantis, *Salmonella* ser Branderup, *Salmonella* ser Paratyphi, *Salmonella* ser Worthington, *Salmonella* ser Give and *Salmonella* ser Dublin. The minimum concentration of Micro-Aid[®] (MIC) required to inhibit the growth of each serotype was determined. This testing procedure has consistently provided quantitative results when Micro-Aid[®] has been tested against a variety of different bacterial isolates, both Grampositive and Gram-negative, aerobic and anaerobic.

Following establishment of the MIC, a concentration-dependent killing assay (time-kill assay) was conducted against each serotype. The time-kill assays involved exposing broth cultures of each of the six *Salmonella* serotypes to Micro-Aid[®] concentrations that were 0, 0.5x, 1x, 2x, and 4x the MIC for each strain tested. Bacterial growth was monitored at hourly intervals over the six-hour time-course of the study. From data generated from the time-kill assays the impact Micro-Aid had on bacterial growth was assessed.

Results

All *Salmonella* serotypes tested were inhibited by Micro-Aid[®] concentrations of 10 mg (MIC) of saponin per mL of media (Table 1). These MICS are comparable to the MICs for many other species of bacteria, e.g., *E. coli, Clostridia perfringens,* etc., that have been tested to date.

Figure 1 depicts the time-kill assay results for *Salmonella* Worthington and the effect exposure to Micro-Aid[®] at increasing concentrations had on bacterial growth over a six-hour incubation period. The

Table 1. Minimum Inhibitory Concentration (MIC) of Salmonella Bacteria Exposed to Micro-Aid®

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Species	MIC (mg of Saponin/mL)		
Salmonella Infantis	10		
Salmonella Branderup	10		
Salmonella Paratyphi	10		
Salmonella Worthington	10		
Salmonella Give	10		
Salmonella Dublin	10		

conditions under which each growth curve was generated was the same, i.e., medium, inculum, incubation temperature and time. The only difference was the presence of Micro-Aid[®] at concentrations of 5 mg/ml (0.5x MIC)

¹ This research was conducted by Dr. R. D. Walker of Anti-infectives Research Consultants (ARC), LLC.

Micro-Aid" in all feed, all the time

RESEARCH SUMMARY

10 mg/ml (1.0x MIC), 20 (2x MIC) and 50 mg/ml (4x MIC). Although only the results for Salmonella Worthington are presented. there were consistent similarities in results among all six Salmonella serotypes tested: 1) The increase in viable cell counts in each control suspension (no Micro-Aid[®]); 2) the patterns of growth of those suspensions exposed to Micro-Aid[®] at 0.5x their MICs were like the Controls; 3) the suspensions incubated with Micro-Aid[®] at 1x their MICs exhibited a bacteriostatic growth pattern; 4) growth pattern of those bacterial suspensions incubated with Micro-Aid® at concentrations that were 2x their



MICs were mixed. Some appeared static whereas others died out during the six-hour incubation time; and 5) the elimination of viable bacteria in all suspensions containing Micro-Aid[®] at 4x the MICs of the bacteria in that environment.

To assess the impact Micro-Aid[®] had on the growth of this bacterium in vitro we compared the percent increase in bacterial numbers for each Micro-Aid[®] concentration the bacterium was exposed to. While the results among all six serotypes was similar we present here the data for *S*. Worthington (Table 2). For the Control suspension, there was a 59.02% increase in viable cell numbers during the four-hour interval between the sample collected at 1 hour and the sample collected at 5 hours of incubation. For the bacteria incubated with Micro-Aid[®] at 0.5x the MIC, there was 51.53% increase in viable cell numbers. The cell count was further reduced to a 4.51% increase for those exposed to Micro-Aid[®] at 1x the MIC. For the bacteria incubated with Micro-Aid[®] at 2x and 4x the MIC, there was bacteria recovered after a four-hour and two-hour exposure, respectively.

This data clearly shows that Micro-Aid[®] has an adverse effect on susceptible bacteria. It is not clear if this is due to Micro-Aid[®]'s impact on the growth rate of the bacteria or if Micro-Aid[®] is bactericidal to susceptible bacteria.

This data clearly shows that Micro-Aid[®] has an adverse effect When Incubated with Micro-Aid[®]

	Control	0.5x MIC	1x MIC	2x MIC	4x MIC
Concentration 1 hr	5.491	5.063	5.322	5.230	4.176
Concentration 5 hr	8.732	7.671	5.562	0.000	0.000
Difference	3.241	2.609	0.240	-5.230	-4.176
% Change 1 vs. 5 hr	59.02	51.53	4.51		

Key Summary Points

- Salmonella were all found to have a similar minimum inhibitory concentration to Micro-Aid[®].
- Although there are over 2,500 different Salmonella serotypes, the impact Micro-Aid[®] had on these isolates and the consistency of their response of this dataset can translate to Salmonella from other sources.
- ✤ Interestingly, this genus of bacteria was comparable in susceptibility to Micro-Aid[®] as all other members of the *Enterobacteriaceae* (e.g., *E. coli*) that have been tested to date.

